

CELLION
Workshop Univ. Surrey / GCI
April 2004

Track visualisation

GSI Material Sciences

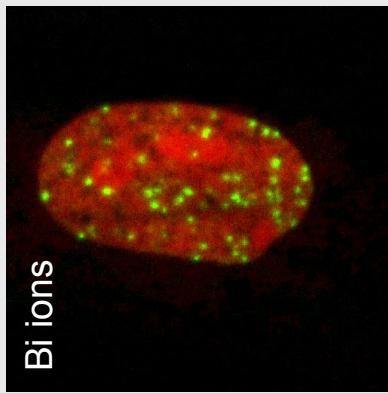
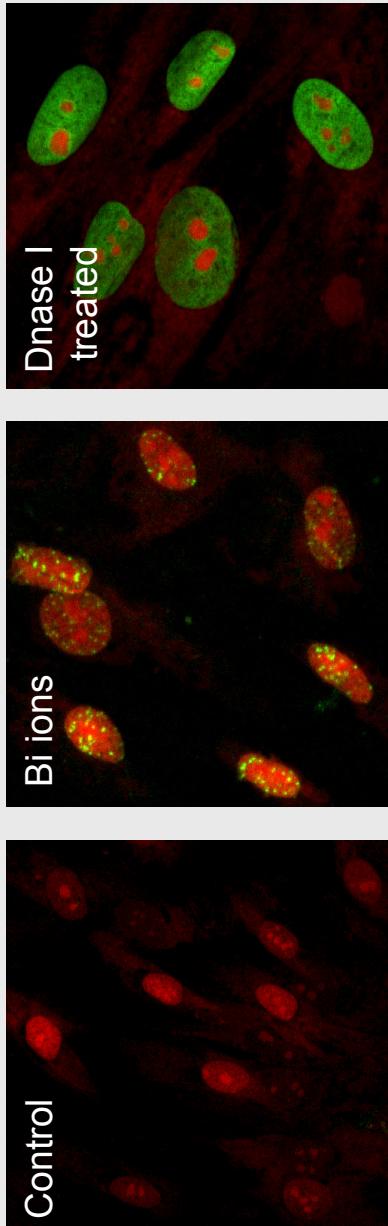
B. Fischer
M. Heiß

GSI Biophysics

G. Becker
C. Fournier
B. Jakob
G. Taucher-Scholz

The basis of subnuclear track visualisation

Particle irradiation: localized deposition of dose
localized production of DNA damage



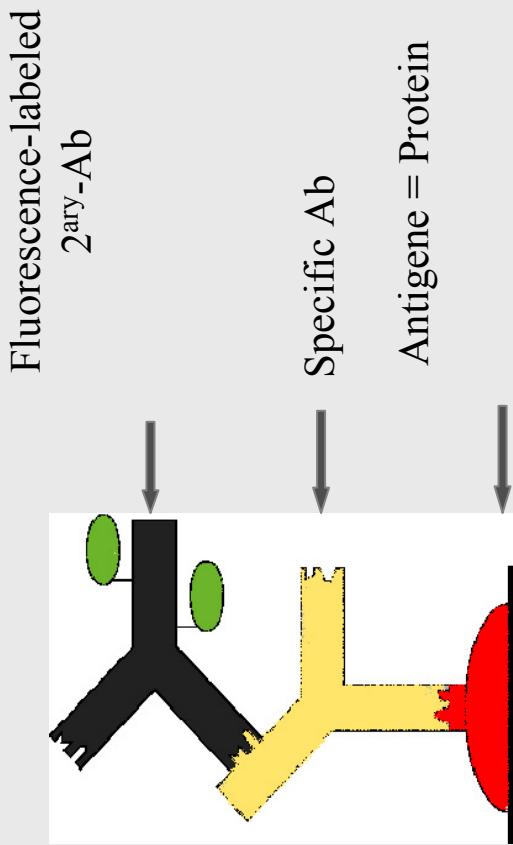
Visualization of the
localized production of DNA **DSBs**
after Bi ion irradiation
and TUNEL assay (FITC-dUTTP)

Disadvantages: low sensitivity
stringent conditions

4.1 MeV/u Bi, 2×10^7 P cm⁻²

Track visualisation

Immunocytochemical detection of proteins recruited to sites of damage



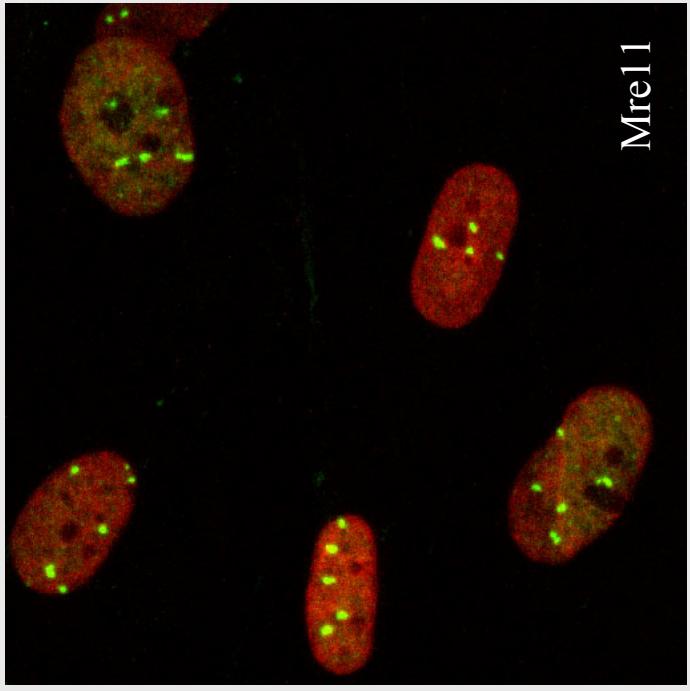
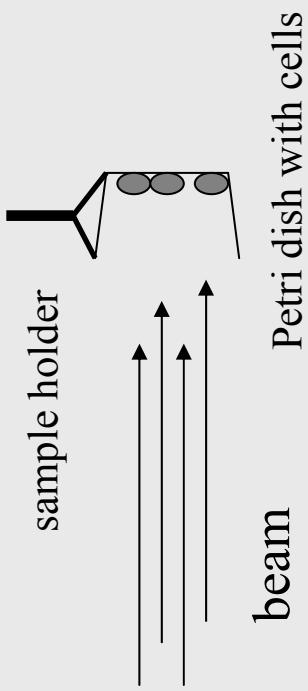
- Cell permeabilisation
- Fixation
- Immunostaining
- DNA-counterstaining
- (confocal laser) microscopy

Requirements:

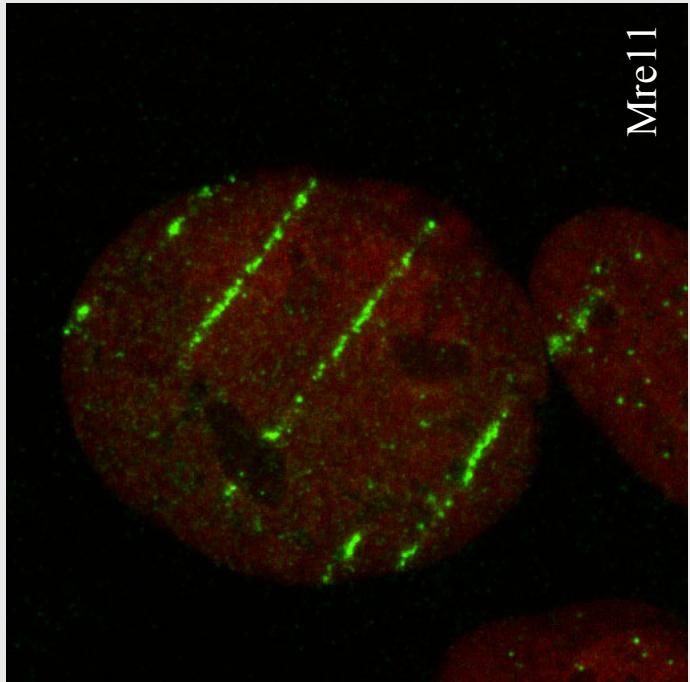
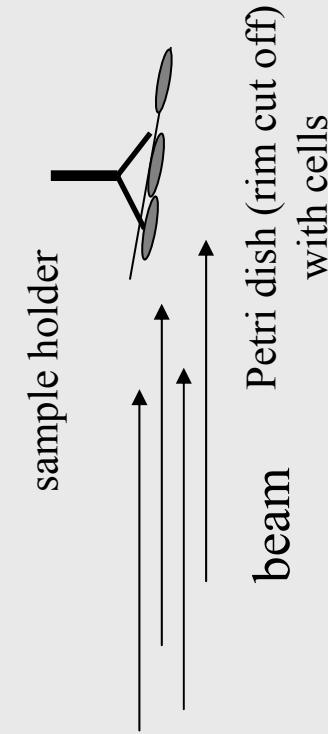
- Rapid concentration of proteins at damaged sites
- Specific response to IIR-induced DNA damage
- Response in all cells (independent of cell cycle phase and cell status)

Biological visualization of single particle tracks (Mre11)

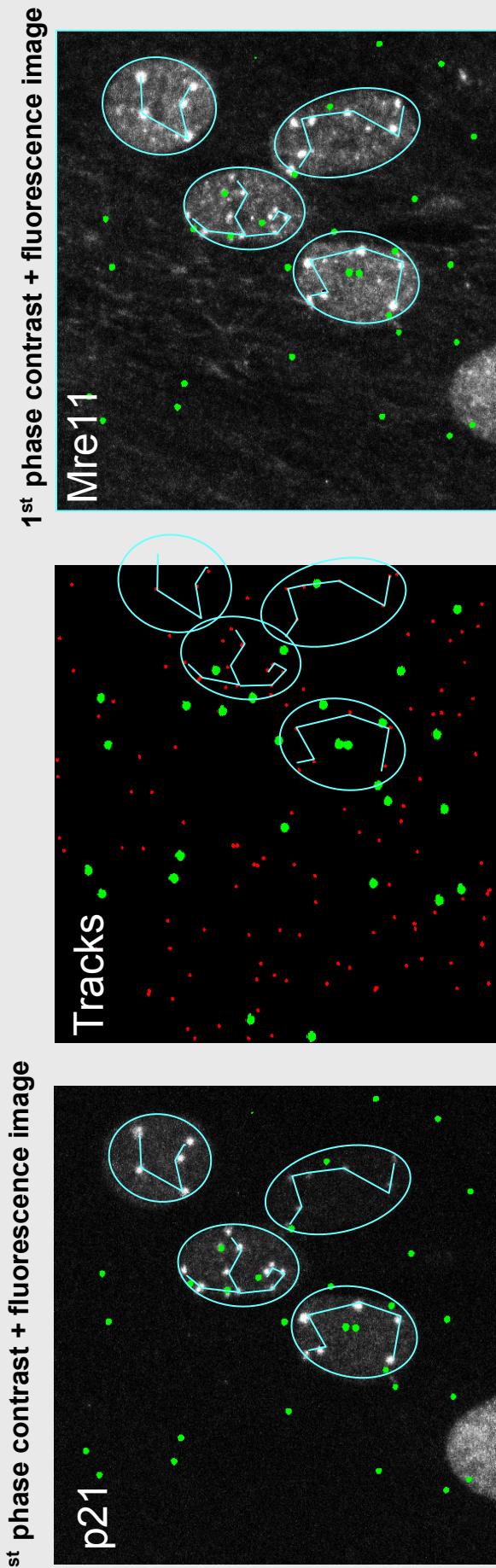
irradiation perpendicular to cell monolayer



irradiation under a small angle ($< 5^\circ$)



Correlation of protein foci with ion tracks



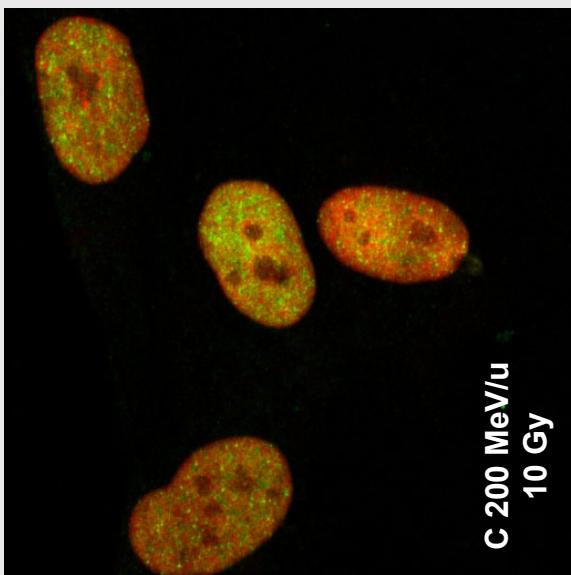
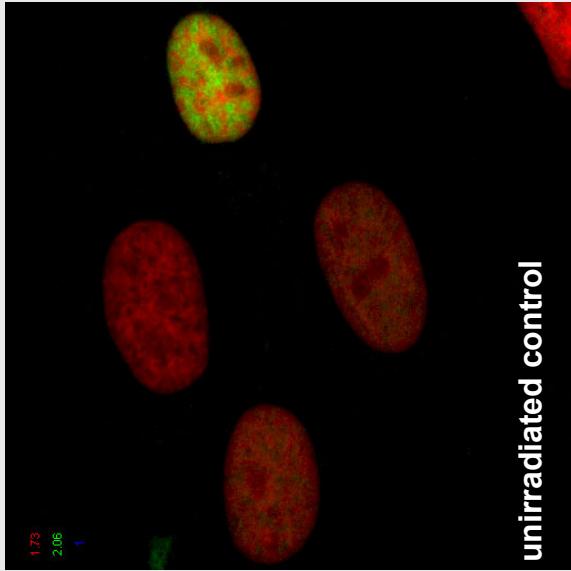
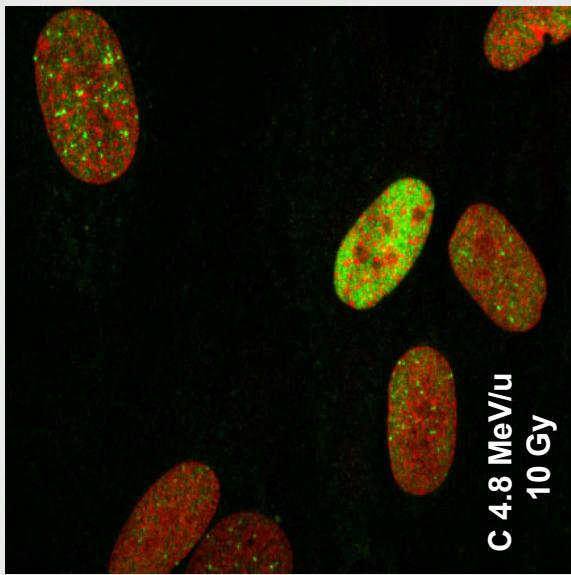
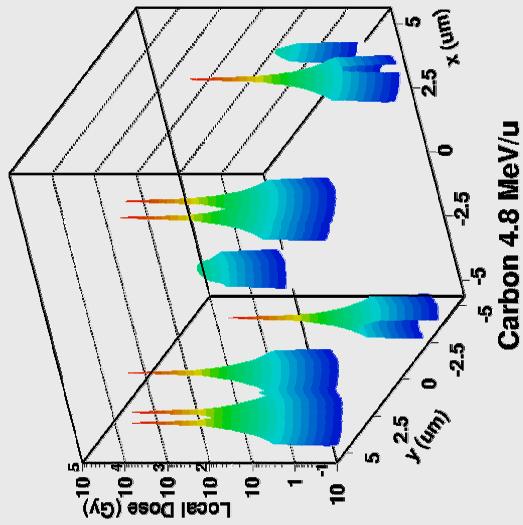
Green: reference tracks
Red: nickel irradiation tracks

- p21:
 - each focus corresponds to an irradiation track
 - not all cells show foci (e.g. S-phase)

- Mre11:
 - all cells show foci
 - each irradiation track leads to a focus
 - additional foci may be detected (e.g. replication sites)

The pattern of immunofluorescence corresponds to the microscopic local dose distribution

High and low energy carbon ion irradiation

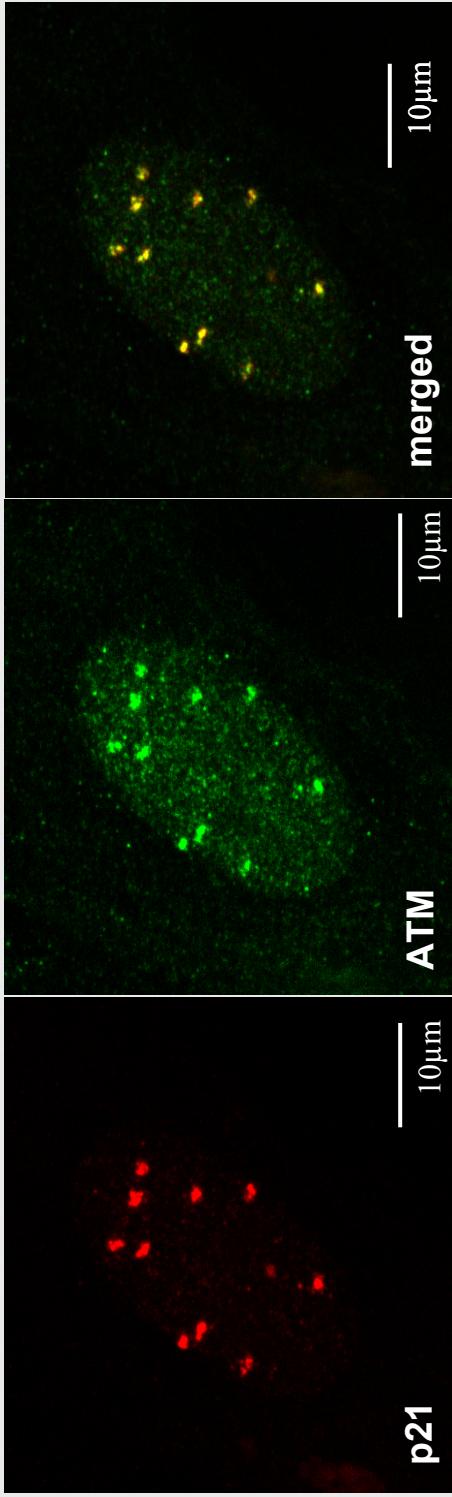


≈300 keV/μm

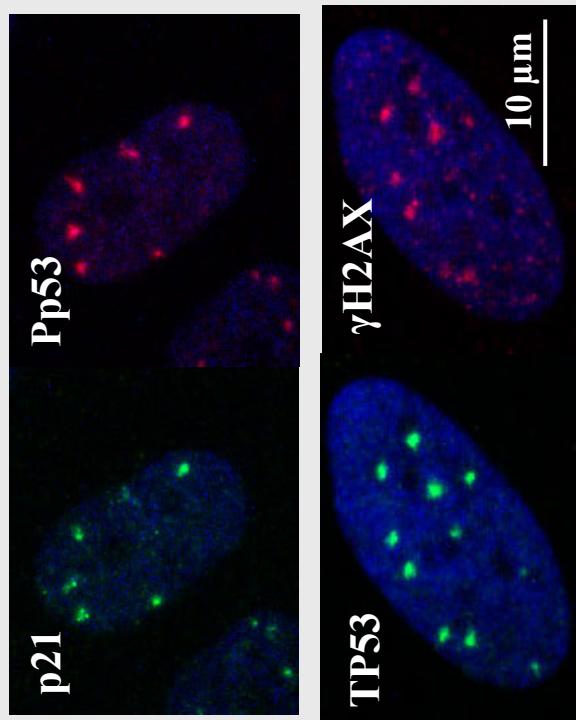
Human fibroblast nuclei immunostained for p21 following exposure to 10 Gy carbon ions

≈16 keV/μm

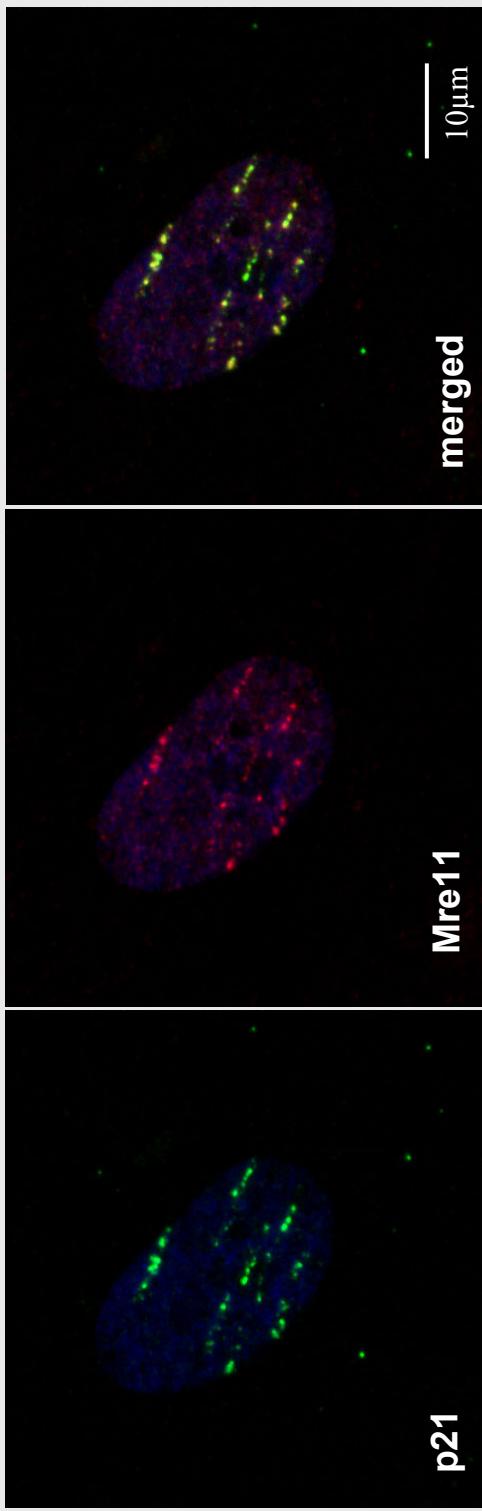
Co-localization of various proteins in foci at sites of ion traversal



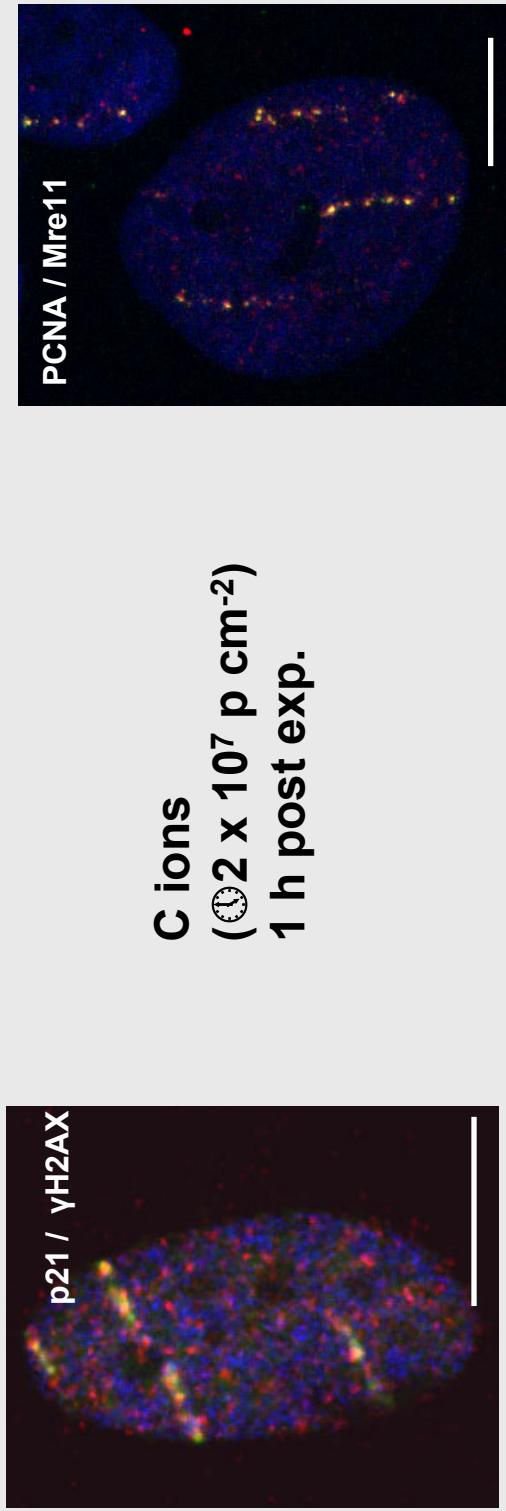
Colocalization of p21 and ATM to foci in nuclei of human fibroblasts



Co-localization of various proteins along radiation tracks



Incident beam at low angle: inhomogeneous protein streaks along carbon ion trajectories



C ions
($\odot 2 \times 10^7$ p cm $^{-2}$)
1 h post exp.

Microbeam experiments:

Why are we interested in track visualisation?

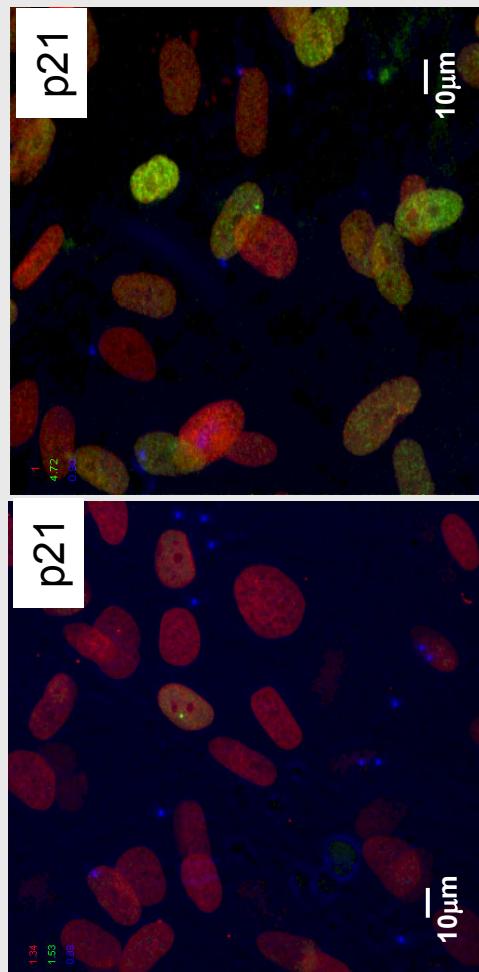
- Biological *in situ* verification of hits (many cells, 1 hit) – nuclear hits
- Estimate of hitting accuracy (many ions /point)
 - Analysis of cellular radiation responses
 - Studies on the response of proteins at damaged sites

Concerns in relation to single track visualisation:

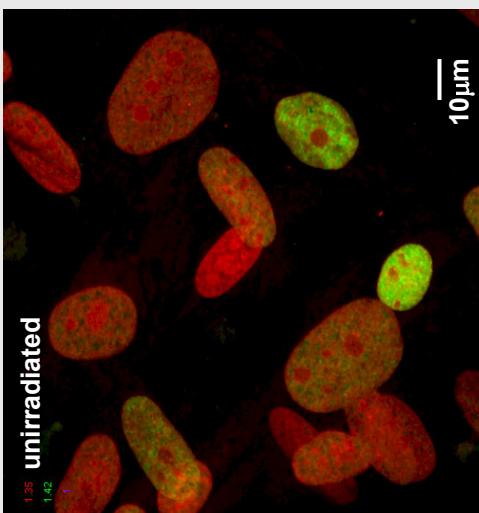
- Sensitivity of foci detection
- Background of foci in unirradiated cells
- Does every particle hit lead to focus formation in the irradiated cell?
 -
- Do all cells respond (independent of cell cycle phase or cell status)
- Do bystander cells respond regarding foci formation?
- Kinetics of foci formation and persistence (postirradiation)

Low fluence irradiation of fibroblasts and retrospective track detection

15 min after irradiation



3 h after irradiation



tracks

Unirradiated control

tracks

Immunostaining:
p21 (green)
PI (red)

Zn 4.9 MeV/u
LET 4000 keV/μm
 $1 \times 10^4 \text{ P/cm}^2$

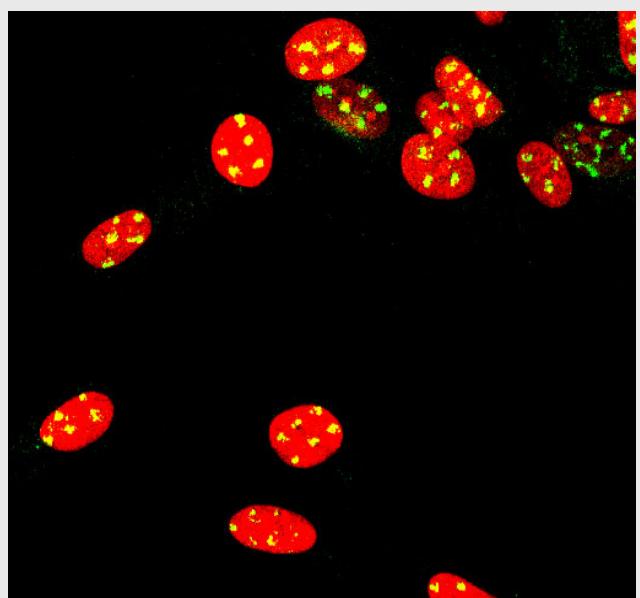
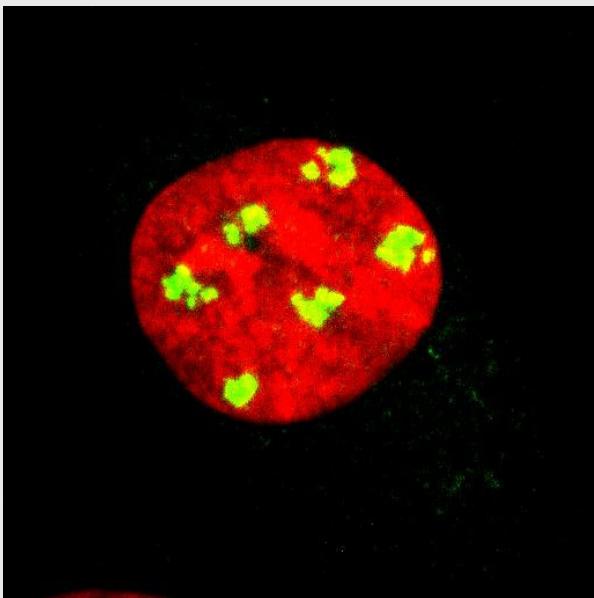
Image size: 160 x 160 μm

2.3 % of nuclei hit

- Cell migration after longer times
- No p21 foci in bystander cells

Microbeam accuracy in cells

Pattern of 53BP1 foci
in fibroblasts nuclei after irradiation
with regularly spaced C ions ($7 \times 7 \mu\text{m}$)
4.8 MeV/u



Rotation of cells
after irradiation

Track visualisation: SNAKE (Munich), RAD51 staining

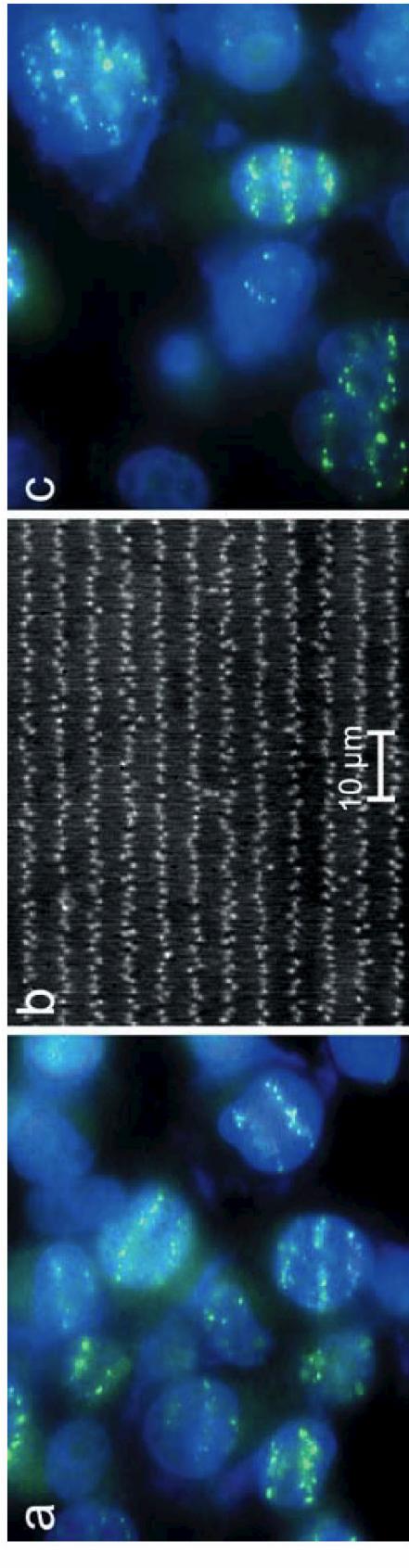


Fig. 8 Immunofluorescence micrograph of cells from irradiation sub-pattern III, with Rad51 foci depicted in green and DAPI staining of cell nuclei in blue **a, c**. For comparison, a cut-out of the nuclear track detector, which was irradiated with the same pattern, is shown at the same magnification **b**. The scalebar is valid for all panels. Note an example for in-plane rotation in panel **a**, and shifted line pattern in the lower section of panel **c**

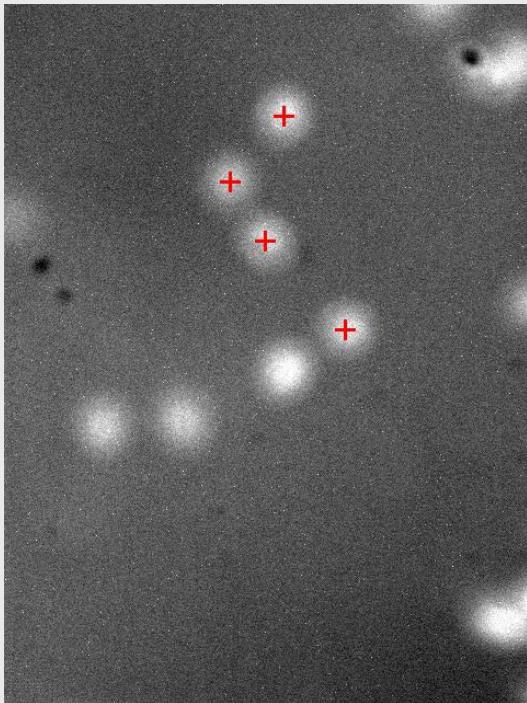
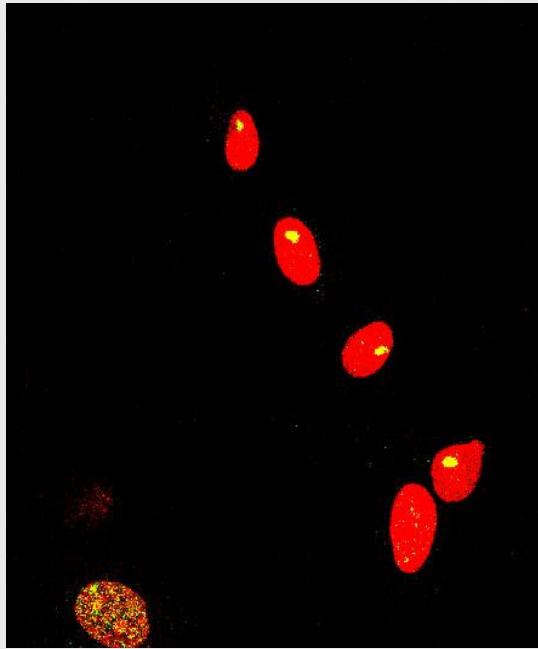
Hauptner et al., Radiat. Environ. Biophys., 2004

**HeLa cells irradiated with 100 MeV O ions
1 μm lateral / 5 μm between lines
1 h postirradiation incubation**

Targeting of single cells with defined number of ions

Irradiation:
carbon ions (4.8 MeV/u)

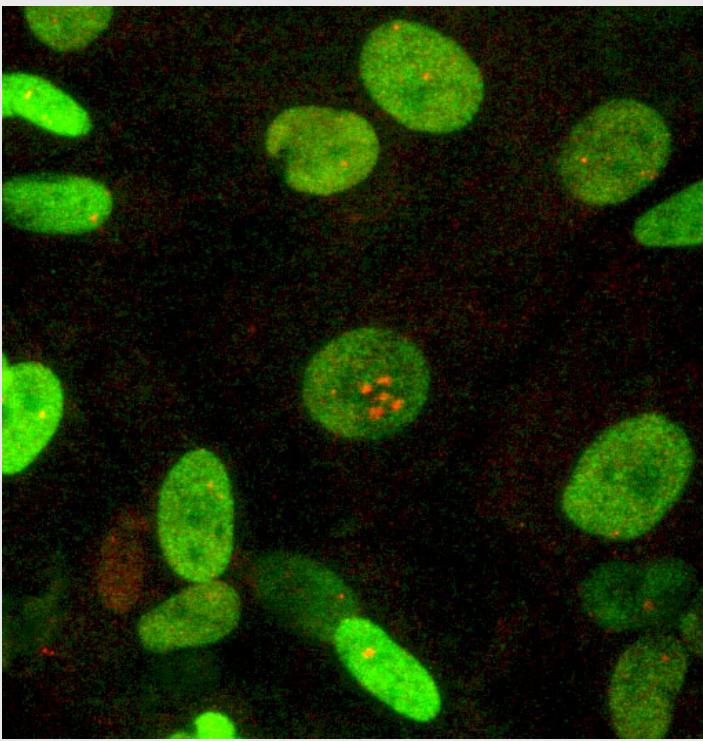
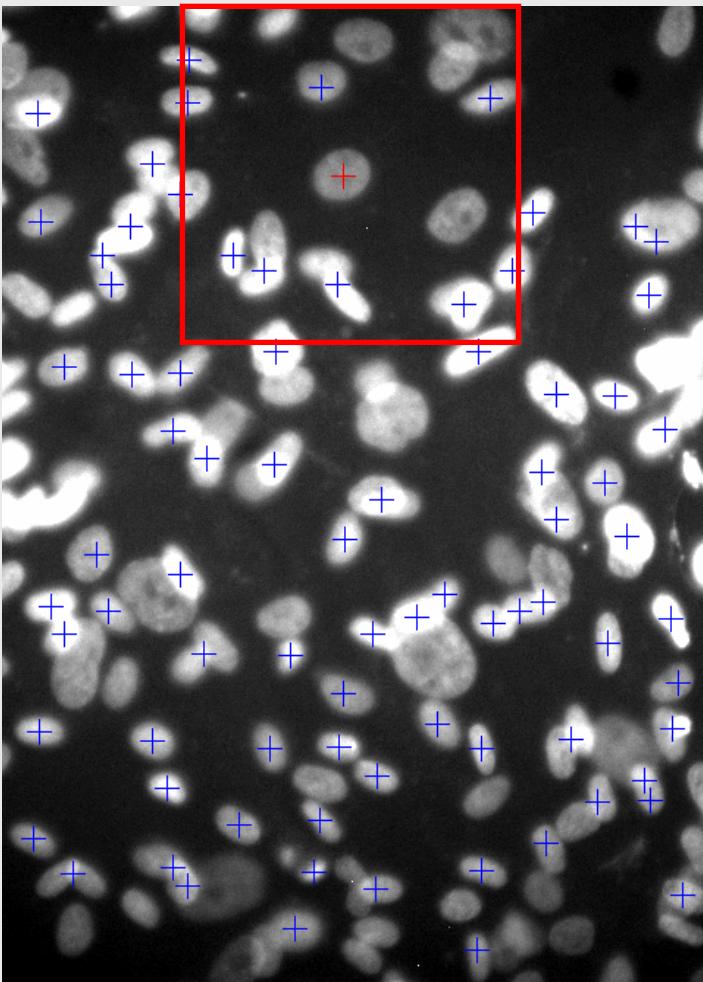
Detected and targeted cells in irradiated fibroblast nuclei
53BP1 foci



Cells targeted after cell detection
and irradiated with 16 C ions per spot
53BP1 foci
after 1h immunostaining and
confocal microscopy
(notice cell migration)

Targeted irradiation of single cells

**with 5 hits cross of 3 spots/axis
5 ions each at 3 μ m distance**



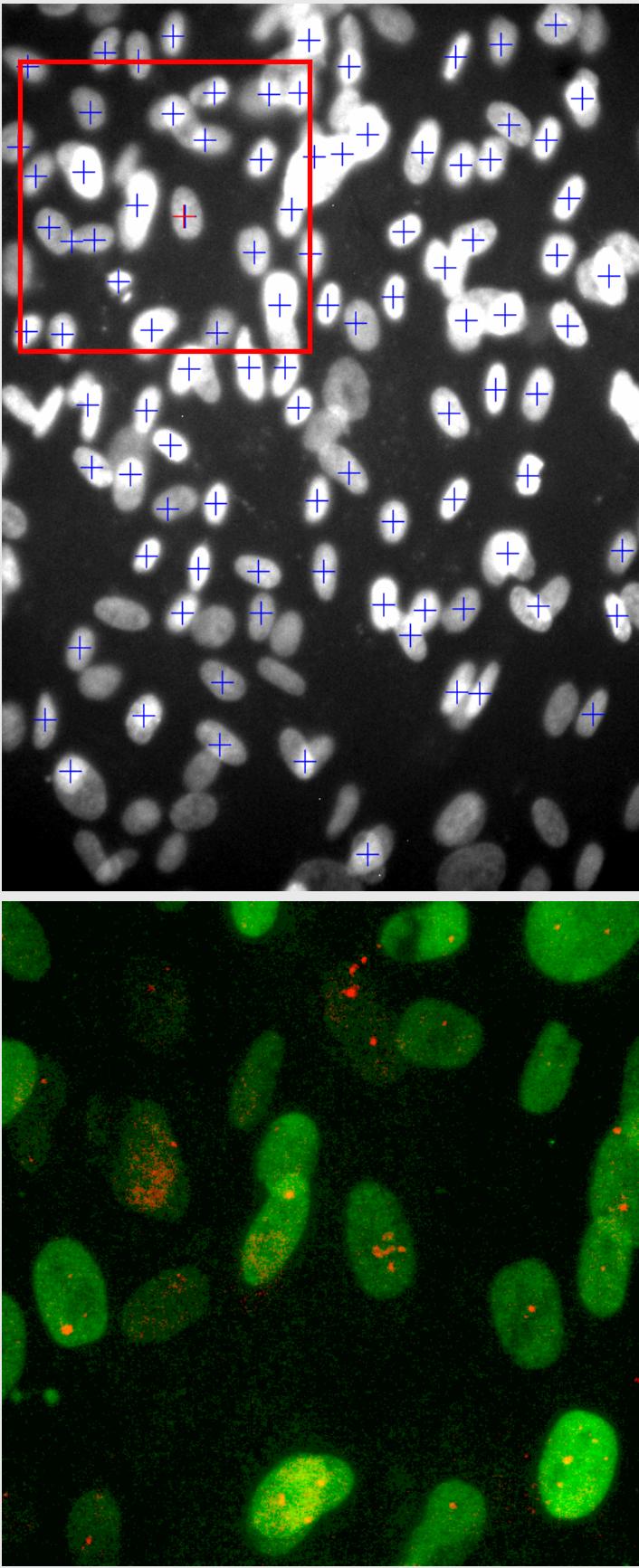
**Immunostaining p21 / 53BP1
1h post irrad**

Very little no. of foci in non-hit cells

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Targeted irradiation of single cells

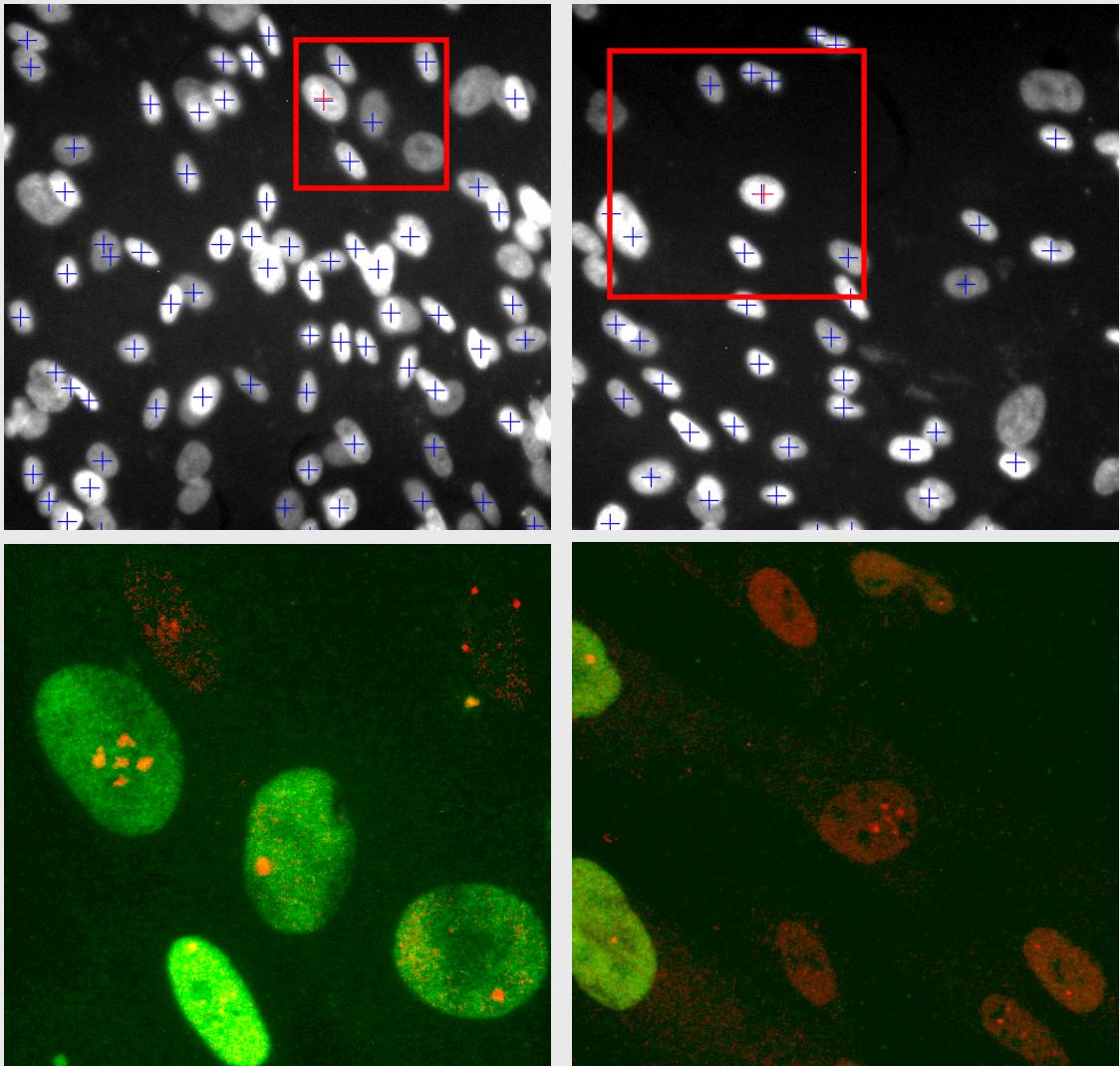
**with 5 hits cross of 3 spots/axis
5 ions each at 3 μ m distance**



**Immunostaining p21 / 53BP1
1h post irrad**

Considerable no. of foci in non-hit cells
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Heterogeneous response of single irradiated cells



Single cell irradiation
with carbon ions

cross of 5 spots
5 ions each
3 μ m distance

p21 / 53BP1
1h post irrad.

Summary

Development phase of **biology** and first **microbeam** experiments

- Cells can be grown to as single cells or to confluence in microbeam chambers.
- Hoechst and UV for cell detection are tolerated under the conditions applied.
- A regular pattern of ion traversals can be reproduced as a biological response.
- Defined single cells can be targeted with a determined number of heavy ions.
- **Track visualization is a useful tool to verify hit detection even after several hours post-irradiation incubation, but critical factors need to be considered.**
- Ongoing biological experiments (p21 induction in bystander cells).